The effect of Methylene Blue in combination with red visible light on model viruses inactivation and coagulation factors in fresh frozen plasma

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Abstract

Background and Aims: Fresh Frozen Plasma (FFP) is one of blood components. The risk of transmission of viruses from blood components regardless selection of blood donors and screening donated blood still remains. There are several methods for viral inactivation. In this study methylene blue(MB) photo inactivation process was used for inactivating viruses

Materials and Methods: In this study Methylene Blue(MB) was used in final concentration of 1µM. Infected Fresh Frozen Plasma (FFP) illuminated by 143 Pieces (PCs) of 1 w red Light Emitting Diodes (LEDs) from two side for 10,15 and 30 minutes and shacked 30 cycle in minutes. the central wavelength of these LED is 627 nm with 20 nm Full Width at Half Maximum (FWHM).Herpes simplex virus-1(HSV-1) and vesicular stomatitis virus(VSV) were used as model viruses to evaluated illumination effects on viral inactivation. level of fresh frozen plasma (FFP) coagulation factors such as fibrinogen, FV, FVIII, protein C, antitrombin measured pre and post illumination.

Results: Initial HSV-1 and VSV titer were calculated to be 107 and 106.5 TCID50/ml, respectively .The level of viral inactivation was expressed as log-reduction. Titer reduction of HSV for 10, 15 and 30 minutes irradiation with shaking was > 6, ≥ 7 and ≥ 7 log, respectively. the ratio of coagulation factors activity remaining unchanged after pathogen inactivation with MB calculated. illumination had a major effects on the mean levels of fibrinogen and FVIII. Significant differences between level of factors before and after illumination were evaluated with a t test for paired samples. No significant differences were seen in the FFP coagulation factors before and after illumination.(P˃0.05)

Conclusions: As results show the optimum time for viral inactivation were adjusted to be 15 minutes. Due to the reduction of virus titer at various times, agitation with illumination is effective.

Keywords: methylene –blue, light, virus inactivation, shacking.

Introduction

Viral contamination of biological products is of concern to public health officials. Pathogen inactivation raise
fresh frozen plasma has been developed in clinical for 15 years(1,2). This system was shown to inactive a broad range of different DNA and RNA viruses in plasma. FFP is used to treat congenital coagulation factor deficiencies, for which specific concentrates are not accessible but Methylene Blue pathogen inactivation process appear to result in loss of coagulation factor leading to additional transfusion of plasma.(2) In this study we systemically investigated the inactivation HSV-1 model for HBV by methylene blue plus visible light treatment while agitate to assess the capacity of this procedure to prevent transfusion mediated HSV transmission. Also level of selected coagulation factors of plasma such as factor V, VIII, antitrombin activity, protein C and fibrinogen before and after photoinactivation evaluated.

**Methods**

**Fresh Frozen Plasma (FFP).** Fresh frozen plasma units of O positive blood group got(collected) from Iranian Blood Transfusion Center. This units were free from(were lacked of) HBV, HCV and HIV. All unit of plasma were stored at -70°C.

**Device and Illumination.** Each plasma samples was illuminated by 143 Pieces (PCs) of 1 w red Light Emitting Diodes (LEDs) from two side. These LEDs emit light at central wavelength of 627 nm with 20 nm Full Width at Half Maximum (FWHM). The distance of the middle of the bag from LED arrays, considered to be 4.5cm. This device was designed for light (LED, including the distance and placement of blood bags per LED) with the help of simulation software Wolfram Mathematica® considering the intensity profile LED sand right number of used LEDs. Light emission to the plasma bag was optimized and uniformed.

**Cell culture.** the efficacy of Methylene blue procedure for HSV-1 and VSV inactivation were evaluated using cell-culture-derived herpes simplex virus model virus for HBV and VSV model for enveloped viruses. Both viruses are able to infect selected cell line, including Vero cell line. Vero cell was propagated in Dulbecco's Modified Eagles medium(DMEM), supplemented with 10% fetal bovin serum and penicillin and streptomycin were used in final concentration of 1% then cell culture incubated at 37°C in a humid atmosphere of 5% CO2.

**Virus amplification and titration.** Vero cell line cultured in sterile 96 well plate. To reach confluent cell monolayer cultured plates were infected with HSV-1 and incubated for 1 h at 37°C to allow attachment. cell were washed with phosphate buffered salin(PBS) then DMEM with fetal bovin was added and incubated for 5 day at 37 °C. Cytopathic Effects(CPE) of virus investigated every day and virus titer are expressed as log TCID50 and were calculated according to the method Reed and Munch. Similar work has been done for measure titer of VSV.

**MB/Light treatment.** Fresh frozen plasma unit were MB/light treated. Methylene Blue and HSV were added to plasma.(1cc virus to 10cc plasma), the final concentration of MB is approximately 1µmol/l. Plasma transferred to illumination bag and air was removed from the plasma unit before illumination. Infected samples illuminated from two side for 10, 15 and 30 minutes and shacked 30 cycle in minutes. Before and after treatment a sample taken from the bag to evaluate titer of viruses.

**control groups.** plasma without virus spiking and plasma plus virus without treatment were used as negative control and positive control respectively. To detect of Methylene blue affection on pathogen inactivation one sample of plasma via MB without illumination assessed. Also light without MB was used to measure affection of light solely.

**plasma coagulation factors.** level of some proteins in plasma such as factor V(TriniClot
The effect of Methylene Blue in combination with red visible light on model viruses …

<table>
<thead>
<tr>
<th>Viruses Model</th>
<th>Infected sample before treatment</th>
<th>Infected sample on Vero cell line after treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>HSV-1</td>
<td><img src="image1" alt="Image of HSV-1 infected sample before treatment" /></td>
<td><img src="image2" alt="Image of HSV-1 infected sample on Vero cell line after treatment" /></td>
</tr>
<tr>
<td>VSV</td>
<td><img src="image3" alt="Image of VSV infected sample before treatment" /></td>
<td><img src="image4" alt="Image of VSV infected sample on Vero cell line after treatment" /></td>
</tr>
</tbody>
</table>

Fig. 1. Cytopathic Effects of model viruses pre and post treatment

TM factor V T1505(Tcoag), factor VIII(TriniClot TM factor VIII T 1507), antitrombin activity(STA-STACHOROM AT III REF 00596)(Stago), protein C( STA-STACOL PROTEIN C REF 00747)(Stago) and fibrinogen(TriniCloTM fibrinogen kit T 1301)(Tcoag) were measured pre and post illumination.

Statistics. SPSS 15.00 and t test for paired samples were employed for data analysis.

Results

HSV-1 and VSV titer was calculated to be 107 and 106.5 TCID50/ml, respectively. The level of viral inactivation was expressed as log-reduction. Log reduction of HSV-1 for 10, 15 and 30 minutes irradiation with shaking was≥6.92, ≥7.00 and ≥7.00 log respectively. VSV virus inactivated by 10,15 and 30 minutes illumination. No Cytopathic Effects(CPE) observed after mentioned time.

The ratio of coagulation factor activity remaining after pathogen inactivation with MB appraised. Illumination had a major affects on the mean levels of fibrinogen and FVIII. Significant differences between level of coagulation factors pre and post illumination were evaluated and no significant differences were seen in the factor before and after illumination. (P˃0.05)

As results show, shaking during illuminating is the effective factor on viral inactivation. The best time for viral inactivation was 15 minutes. Due to the reduction of virus titer at various times, agitation with illumination was more effective.

<table>
<thead>
<tr>
<th>Table 1: inactivation model viruses using different illumination time</th>
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<tr>
<td><strong>Model virus(mean titer)</strong></td>
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<tr>
<td>-----------------------------</td>
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<tr>
<td>Exposure time(minutes)</td>
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<tr>
<td>Initial titer</td>
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<tr>
<td>10 min</td>
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<tr>
<td>15 min</td>
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<tr>
<td>30 min</td>
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</table>
Discussion

In present study the inactivation effects of MB/visible light on viruses investigated. Methylene Blue has long been known when it was used to treat malaria(2,13). It used at low doses to treat methemoglobinemia(3,). Methylene Blue can intercalate into DNA or combine with the outer helix, depending on the Mg $^{2+}$ ionic strength and concentration(3). Platelets ,plasma and red blood cells do not require genomic DNA to be viable therefore MB target nucleic acid of pathogen to interrupt their function. beyond pathogen's genetic damage, protein and membrane molecules can be incurred s leading to reduction blood product quality.(3)

In this study, attempted achieve a proper balance between pathogen inactivation and the product quality by MB/visible light method. Also effect of shaking on time exposure investigated.

WHO recommendation show that in the current procedure, individual plasma units are
treated with 1µM methylene blue and white fluorescent light for 1 h at 45000lux or with low-pressure sodium lamps at 200 Joules/cm² for 20 minutes (19). Later, work on other systems (Baxter Pathinact and Maco Pharma Theraflex systems) was performed on fresh plasma (18). Optimum time irradiation of the newest version of the illumination device (Macotronic B; MacoPharma) is 12-15 min for 2 single unit of plasma. This system use visible light (630 nm, 180 J/cm²) and Methylene Blue in 1µm final concentration (12).

In this study optimum light dose 54 J/cm² was calculated. The best time for illumination with the same light dose without shaking to inactive virus was 45 minutes (4). This procedure with shaking reduce this time to 15 minutes.

**Conclusion**

Available data demonstrate that MB/visible light shaking base device is effective on viral inactivation and show quality of coagulation indication factors of FFP during 15 minutes irradiation are better preserved.

**Table 2: Effect of photoinactivation on coagulation factors in FFP**

<table>
<thead>
<tr>
<th>Plasma coagulation factors</th>
<th>Reference</th>
<th>FFP before illumination</th>
<th>10 min illumination</th>
<th>15 min illumination</th>
<th>30 min illumination</th>
<th>Preserve after 10 min illumination (%)</th>
<th>Preserve after 15 min illumination (%)</th>
<th>Preserve after 30 min illumination (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Factor V</td>
<td>60-130</td>
<td>62</td>
<td>65</td>
<td>69</td>
<td>58</td>
<td>100</td>
<td>100</td>
<td>93.5</td>
</tr>
<tr>
<td>Factor VIII</td>
<td>60-150</td>
<td>70</td>
<td>50</td>
<td>44</td>
<td>44</td>
<td>71.4</td>
<td>64.2</td>
<td>62.8</td>
</tr>
<tr>
<td>Fibrinogen</td>
<td>200-400</td>
<td>241</td>
<td>201</td>
<td>188</td>
<td>182</td>
<td>83.4</td>
<td>78.00</td>
<td>75.5</td>
</tr>
<tr>
<td>Protein C</td>
<td>70-130</td>
<td>116</td>
<td>110</td>
<td>108</td>
<td>97</td>
<td>94.8</td>
<td>93.1</td>
<td>83.6</td>
</tr>
<tr>
<td>Antitrombin activity</td>
<td>80-120</td>
<td>109</td>
<td>110</td>
<td>110</td>
<td>108</td>
<td>100</td>
<td>100</td>
<td>99</td>
</tr>
</tbody>
</table>

**References**


