Evidence of H1 and H3 Influenza Virus Infection in Captive Birds in Tehran from November 2008 to February 2009

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Abstract

Background and Aims: Influenza A virus infects birds and some mammalian including human. H1 and H3 subtypes are circulated in both human and birds’ population. To determine the prevalence of the mentioned subtypes in birds, different captive bird species in Tehran zoo, and affiliated centers in Tehran were investigated for virus infection.

Methods: In this study, Between November 2008 and February 2009, 76 cloacal swabs and serum samples were collected from 5 orders of Anseriformes, Galliformes, Columbiformes, Pelicaniformes and Phoenicopteriformes. Antibody surveillance was undertaken by haemagglutination inhibition assay and for detection of influenza virus genome RT-PCR technique was used.

Results: In total, 57.7% and 88.75% of bird sera were seropositive against H1 and H3 viruses respectively. The highest GMT value and the greatest antibody titers were observed in Galliformes order particularly in chicken species. Influenza A genome was not detected in any of the samples by RT-PCR using M gene.

Conclusion: Results of this study indicated that seropositive birds were infected during the last or possibly previous years with H1 and H3 virus strain.

Keywords: Hemagglutination Inhibition Test (HI); Real time polymerase chain reaction (RT-PCR); Captive birds; Tehran

Introduction

Influenza A viruses which belong to Orthomyxoviridae family could infect different avian and mammalian species. The natural reservoir of influenza A viruses are aquatic birds such as Anseriformes and Charadriiformes in which different combinations of all 16 HA and 9 NA has been found (1). Although there has been no report of avian influenza isolation from chickens, turkeys and other Galliformes species in their wild states and they are not natural reservoir of influenza viruses (2), humankind activities such as captivity, domestication and agriculture have changed the natural ecosystem of birds. Zoological collections are man-made systems where captive birds have close contact to free-flying birds, park keeping staff and visitors to parks and zoos. Variety of host bird species in these new niches may alter the ecology and epidemiology of influenza viruses therefore viruses could transmit interspeciesly and be adapted in new hosts (3).
Seroprevalence of H1 and H3 in captive birds…

Although H1N1 and H3N2 viruses are currently circulating among human population (4), but it is believed that all influenza A viruses in mammalians are originated from the avian genes (2). In the 20th century, Asian influenza pandemic of 1957 caused by a reassortant H2N2 virus that some genes of it came from avian viruses and in H3N2 virus of 1968 pandemic (Hong Kong influenza pandemic), the genes of virus originated from reassortment of human and avian genes (5, 6).

In recent years, direct transmission of H5N1 (4, 7), H7N7 (8) and H9N2 (9) viruses from birds to human have been reported. Therefore it is necessary to monitor places such as parks and zoos to track down the influenza A viruses. The present study was undertaken to provide insight about the possibility of influenza H1 and H3 viruses’ infection in captive birds of man-made systems such as zoo and parks where the birds have close contact to human and other influenza viruses’ hosts.

Methods

Sample collection

From November 2008 to February 2009 (during influenza season in Tehran), 76 cloacal swabs and serum samples were collected from birds in Tehran Zoo, Saiee Park and Pardisan Park. Studied birds included, 3 species of Anseriformes order (plus 5 hybrid species of ducks), 7 species of Galliformes order, 1 species of Columbiformes order, 1 species of Pelicaniformes order and 1 species of Phoenicopteriformes order. Cloacal samples using Dacron swabs were transferred into the viral transportation medium (VTM). The VTM was prepared with sterile glycerol-PBS (1:1) solution (pH 7.2) containing Benzyl-Penicillin, Streptomycin and Amphotericin. The samples containing VTM was centrifuged at 3000 x g for 10 min and the supernatants were stored at -20°C for further process. To harvest sera, blood samples were centrifuged at 2500 xg for 15 min and stored at -20°C for further studies.

Viruses

NIBSC standard human influenza A/ New Caledonia/ 20/ 99 (H1N1) and A/ Panama/ 2007/ 99 (H3N2) viruses used in this study were passaged in Madine Derby canine kidney cells (MDCK) in Influenza Unit of the Pasteur Institute of Iran (10).

Vaccination of chickens

Positive control serum for HI assay was not accessible. Therefore, it was decided to immunize seronegative chickens (confirmed with ELISA) with inactivated human influenza vaccine influvac 2008/2009 (Solvay Pharmaceuticals B.V. Weesp, The Netherlands) containing A/Brisbane/59/2007 (H1N1) and A/Brisbane/10/2007 (H3N2) antigens. Group 1, including 4 chickens was vaccinated by injecting 0.25 ml of vaccine (half-dose) containing 7.5 μg of each antigen. Group 2, including 4 chickens, were injected by full-dose vaccine (0.5 ml and 15 μg of each antigen) whereas 2 chickens of group 3 (control group) were not vaccinated against influenza viruses. Vaccination was performed intramuscularly in the breast muscle of birds at day 0 and 21. Chicken sera were collected at days 0 (before first injection), 21 (before second injection) and 35 to obtain antibodies against H1 and H3 viruses.

Serological assay

The Haemagglutination Inhibition (HI) assays were carried out to determine antibody titers against A/New Caledonia/20/99 (H1N1) and A/Panama/2007/99 (H3N2) viruses in serum samples (11). To measure antibody titers, beta procedure of HI test was used (12). The sera of all the species except chickens were pre-treated with 10% chicken red blood cells to remove non-specific hemagglutinating reagents. Also sera of hunting birds (pheasant and partridge), quail and guinea fowl were treated in a water bath at 56°C for 30 minutes. Afterward a 2-fold serial dilution of the treated sera was made in 96-well U shaped plate using PBS (pH 7.2). A 25μl of 4 HA unit antigen was added to each well. The plates were incubated for 30 minutes at room temperature, and finally 1% chicken RBCs suspension (in PBS containing 0.5% v/v bovine serum albumin) added and the plates again incubated for 40 minutes at room temperature. HI end point titers were determined as the reciprocal of the highest dilution that produced complete inhibition of haemagglutinin activity. A titer of 16 and
higher was considered as a positive. Results were validated by negative control serum with titer less than 8, and positive control serum with a definite titer more than 16. In HI assay for each serum sample, RBC control well, serum control well (to control the removal of non-specific haemagglutinating reagents in serum) and virus control well was conducted.

RT-PCR
 RNA extraction from cloacal specimens was performed using RNX-Plus solution (CinnaGen, Tehran, Iran). The procedure was carried out according to the manufacture’s recommendation and RNA used for cDNA synthesis. Briefly, the mixture of 10µl RNA and 2µl Uni12 primer (5′-AGC-AAA-AGG-AG-<G>-3′) was incubated at 70°C for 5 minutes and immediately chilled on ice. Thereafter, the components of RevertAid First Strand cDNA Synthesis Kit (Fermentas, Canada) including 5 x reaction buffer (4µl), Ribolock TM RNase Inhibitor 20U/ml (1µl), dNTP mix 10mM (2µl) and ReverAidTM M-MuLV Reverse Transcriptase 200u/µl (1µl) was added to the tube containing RNA and primer. The tube was incubated for 5 minutes at 25°C followed by 42°C for 60 minutes and finally at 70°C for 5 minutes. M primers targeting matrix gene of influenza type A viruses were used in PCR (MF: 5′–GACTCAATGTCAAGAACCTTTA–3, MR: 5′–CCACTTATTTCTCTGTGTTAG–3′) to amplify a 132bp fragment. The PCR reaction mixture consisted of 6µl cDNA, 12.5µl Taq DNA Polymerase Master Mix RED kit (Ampliqon, Denmark) which contained Tris-HCl 150mM pH 8.5, (NH4)2SO4 40mM, MgCl2 1.5mM, 0.2% Tween 20, dNTPs 0.4mM, Ampliqon Taq DNA Polymerase (0.05Unit/ul), 1µl of each forward and reverse of M-primers (Table 1) and distilled water to the final volume of 25μl. The thermocycling conditions were 95°C for 5 minutes (primary denaturation), 35 cycles of 94°C for 40 seconds (denaturation), 59°C for 40 seconds (annealing) and 72°C for 40 seconds (extension), and the final extension at 72°C for 5 minutes which terminated the PCR reaction. The PCR products were analyzed by running 2% agarose gel electrophoresis.

Sensitivity of RT-PCR
 The sensitivity of RT-PCR was assessed by ten-fold serial dilution of extracted H1N1 RNA (A/ New Caledonia/ 20/ 99). Subsequently, RT-PCR was performed according to the procedure described above. The highest dilution of RNA with positive RT-PCR was considered as the sensitivity cut-off.

Results
Vaccination of chickens
 The serologic results of vaccinated chickens are summarized in Table 1. Mean titer of HI antibodies against human vaccine antigens in full-dose vaccinated chickens were higher than half-dose vaccinated (4 fold for H1 and 2 fold for H3 antibodies). Control group showed no antibody titers in HI assays. Prevaccination chicken sera (day 0) were seronegative for human antibodies. Antibody mean titers against vaccine antigens at day 35 were significantly higher than those at day 21. First vaccine injection resulted in H3 antibody titer production in 75% and 100% of half-dose and full-dose vaccinated chicken respectively, whereas H1 antibody was not detected in any of the sera. At day 35 (2 weeks after second injection), 34% of half-dose vaccinated chickens had H1 antibody titer and 100% showed antibody against H3 antigen. Group 2 (full-dose vaccinated) had 100% seropositivity for each vaccine antigens. Positive serum resulted from vaccination was used as control serum in HI assay.

Serological assay
 In total, 57.7% (41 of 71) of bird sera were positive for antibodies against H1 subtype (HI titer 1:16≤ was considered as positive) and also 62 of the 70 birds’ sera were positive for H3 subtype antibody with overall percentage of 88.75%.

HI results for H1 subtype
 Results of HI assay against H1 subtype among different bird species is shown in Table 2.
Seroprevalence of H1 and H3 in captive birds …

Table 1. The results of chicken vaccination.

<table>
<thead>
<tr>
<th>Day Post-Vaccination</th>
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<th>35</th>
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<td>Antigen Subtypes</td>
<td>H1</td>
<td>H3</td>
<td>H1</td>
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<td>Half-dose vaccinated Chickens</td>
<td>0%*</td>
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<td>(MT: 16)</td>
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<td>Full-dose vaccinated Chickens</td>
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<td>(MT: 16)</td>
<td>(MT: 64)</td>
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<td>Unvaccinated Chickens</td>
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- MT: Mean Titer is the average titer of H1 in each group.
- %: % represents the percentage of positive serum samples in each group.
- Vaccination was carried out at day 0 and 21. Sera collection was at day 0, 21 and 35.

Total Geometric Mean Titer (GMT) value of H1 antibody titer was 16 with the antibody range of 2 to 256. As shown Galliformes had the greatest GMT value of 30.33 while that in Anseriformes was 9.07. Phoenicopteriformes came afterward with the GMT value of 8 followed by Columbiformes (3.62) whereas the only sample of Pelicaniformes had antibody titer of 8.

Of Anseriformes order, Geese species had GMT of 10.55 which was higher than Ducks species with GMT of 8.72. In serum of Swan sample no antibody titer against H1 subtype was detected. In Galliformes order, the highest

Fig. 1. Percentage of seropositivity against H1 and H3 subtypes in different bird orders.

Fig. 2. Contribution of different bird species in seropositivity of Anseriformes order.

Fig. 3. Contribution of different bird species in seropositivity of Galliformes order.

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<table>
<thead>
<tr>
<th>Order/Species</th>
<th>No. of Samples</th>
<th>No. of Positive Samples</th>
<th>% of Positive Samples</th>
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<td>Phoenicopteriformes</td>
<td>8</td>
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**Table 2:** Distribution of Antibody Titers against Avian Influenza H1 Subtype Using Hemagglutination Inhibition Assay
Table 3: Distribution of Antibody Titers against Avian Influenza H3 Subtype Using Hemagglutination Inhibition Assay

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<thead>
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<th>1:8</th>
<th>1:16</th>
<th>1:32</th>
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GMT value of H1 antibody was found in chickens (73.51) followed by Partridges (24.67), Pheasants and Quails (22.62), Guinea fowls (13.45) and Turkeys (11.31). Figure 1 represents data on the distribution of H1 antibody among the birds’ sera. As shown 57.7% (41/71) of serum samples had antibody. Galliformes had the highest seropositivity prevalence (33/39, 84.61%) against H1 subtype whilst 31.81% (7/22) of Anseriformes and 50% (1/2) of Phoenicopteriformes were found seropositive. In Columbiformes and Pelicaniformes no antibody titer of H1 subtype was detected. Figure 2 demonstrates the contribution of each bird species in seropositivity of Anseriformes order against H1 subtype. Ducks had the seropositivity of 71.42% (5/7) whereas the seropositivity of Geese was 28.57% (2/7). As illustrated in figure 3, the highest contribution of seropositivity in Galliformes belonged to chicken species (45.45%, 15/33) followed by 24% (8/33) in Partridge species, 15.15% (5/33) in Guinea fowl species, 6% (2/33) in Turkey and Quail species and 3% (1/33) in Pheasant species.

**H1 results for H3 subtype**

Out of 70 serum samples collected from different bird species, 62 samples showed antibody titer against H3 subtype with total GMT value of 33.29 in antibody range of 4 to 256. Galliformes had the highest GMT value of 57.36 (Table 3) following by Anseriformes (19.94), Phoenicopteriformes (16) and Columbiformes (14.49). The sample of Pelicaniformes had no antibody titer against H3 subtype (titer of 8).

Analysis of data implied that in Anseriformes order, GMT value in Geese was 21.11 in comparison with the GMT value of 18.22 in Ducks. Swan serum had no antibody titer against H3 subtype. In Galliformes order, the greatest GMT value was in chicken species (90.50) whereas GMT value was 64 in Quail species, 58.68 in Partridge species, 45.25 in Guinea fowl and Pheasant species and 19.02 in Turkey species. Prevalence of antibody titer against H3 subtype in 70 birds was 88.57% (Figure 1). The highest seropositivity rate of 100% was in Galliformes (38/38). Anseriformes showed H3 antibody seropositivity of 86.36% (19/22). It was seen that 57.14% (4/7) of Columbiformes and 50% (1/2) of Phoenicopteriformes were seropositive for H3 antibody in serum samples. Pelican serum was not seropositive for H3 subtype antibody.

Figure 2 represents the distribution of bird species against H3 subtype in Anseriformes: Seropositivity for H3 antibody was 68.42% (13/19) in Ducks, 26.31% (5/19) in Geese and 5.2% (1/19) in Swans. The highest seropositivity in Galliformes order was related to chickens (36.84%, 14/38) followed by 21.05% (8/38) in Guinea fowls and Partridges, 10.52% (4/38) in Turkeys and 5.26% (2/38) in Pheasants and Quails (Figure 3).

In tested birds of Pardisan Park, 44.4% (4/9) of serum samples had H1 antibody whereas H3 antibody was observed in all 9 sera. Thirteen of the 30 (43.3%) birds’ sera of Saiee Park had antibody titers against H1 subtype as compared to 25 of the 29 tested sera (86.2%) that showed titers for H3 antibody. Of 32 birds sera belonged to Tehran Zoo, 24 (75%) were positive for H1 antibody and 28 (87.5%) had antibody titers against H3 subtype.

**Molecular assay**

The sensitivity of RT-PCR was determined to be 0.1ng template RNA genome. No avian influenza viruses’ genome was detected in any of 76 cloacal samples collected from Tehran Zoo, Saiee Park and Pardissan Park in Tehran between November 2008 and February 2009.

**Discussion**

To assess the prevalence of H1 and H3 viruses in captive birds of Tehran, cloacal swabs and serum samples were collected from Tehran Zoo, Saiee Park and Pardissan Park. Our study is one of the rare surveillance of H1 and H3 influenza viruses among a variety of captive bird species in Iran. Studied sites had the importance in the ecology of influenza viruses due to the variety of captive bird species and their close contact with the free flying birds and human. Furthermore, in Tehran Zoo, some influenza hosts such as horse and dog were kept. Diversity of bird species in the studied
zoo and parks could lead to a unique condition that facilitates mutations and altering of viruses’ ecology to adapt to new hosts including other bird species and human. Haemagglutination inhibition assay is routinely used to detect serum antibodies of influenza A viruses. The ability of influenza viruses to agglutinate erythrocytes is related to receptors specificity. It has been shown that avian influenza viruses bind preferentially to the N-acetylneuraminic acid 2, 3-galactose (NeuAc2, 3-Gal) linkage, whereas human viruses prefer the NeuAc2, 6-Gal linkage (13). Chicken red blood cells contain both NeuAc2, 3-Gal and NeuAc2, 6-Gal receptors so they can be used in HI assay.

RT-PCR is a valid and universal molecular technique to detect and confirm the presence of influenza viruses’ genome even if they are present at a very low level in the laboratory specimens. The primers used to detect influenza type A viruses, were designed from highly conserved regions of M gene and were type specific which could detect M segment of influenza type A viruses with origin of various hosts (14). This assay is more sensitive, more specific and less time consuming in comparison with other diagnostic assays (15). However, the absence of the expected RT-PCR products, i.e. a negative result, does not necessarily mean the absence of influenza A viruses. Results should be interpreted along with information about clinical signs and epidemiological data.

Interestingly, during our study period, there was no report of death or clinical signs of influenza disease among the captive birds, whereas the birds were not vaccinated already against influenza A viruses. Although the results could be attributed at least partially to the presence of an undetectable amount of genomic RNA, based upon the sensitivity of the test, our findings demonstrated that no RNA genome of influenza A viruses was present in the samples under study. Despite H1 and H3 are most currently subtypes in human population and there is limited information on the prevalence of these two subtypes among birds, but some previous studies have been reported presence of antibodies to H1N1 and H3N2 viruses in wild and domestic birds in Brazil (16). Our study showed a high seropositivity of 57.7% against H1 subtype and also 88.75% against H3 subtype. Galliformes order had the highest seropositivity and antibody titer among tested birds. The present study is supported by other studies reporting that H1 is one of the predominant subtypes among domestic poultry (3). Meanwhile, ability of H3 virus to infect of chickens and turkeys is proved by previous studies (17).

Among all studied birds species, chicken showed the highest seropositivity and GMT value against influenza viruses. In Hong Kong, H1 antibodies to A/HKU/10/77 (H1N1) human virus was detected in 19.4% of examined hen sera (18). Ayoub et al (19) reported a positive H1 antibody titer against human H3N2 viruses in hens and turkeys. In hens, 43- 64.7% were seropositive whereas 70.7- 78% of turkeys had antibodies against human H3N2 viruses. Seropositivity of chickens is noteworthy because this species is considered as a popular domestic poultry especially in rural areas that have close relationship to human.

The results of our study represented seropositivity for H1 and H3 viruses in pheasant species but low GMT value occurrences. Previous studies implied that in infection of pheasant by influenza viruses, low levels of serum hemagglutination inhibition antibody titers would be detected when chicken red blood cells were used in HI assay. More importantly, pheasants can serve as a "carrier" of influenza A viruses because of their continuous asymptomatic infection (20).

On the other hand, previous studies indicate that quails play a significant role in the interspecies transmission of influenza virus from ducks to chickens (21). Domestic poultry such as quail and chicken possess α2, 3 and α2, 6 sialic acid receptors that may help passage of avian influenza viruses from bird species (such as ducks with only α2, 3 sialic acid receptor) to human with predominant α2, 6 sialic acid receptors (22, 23). As sample collection of this study was undertaken in an environment that no pigs were kept, chicken and quail may have
a key role in the transmission of influenza virus from birds to human and vice versa. *Anseriformes* order especially ducks and geese showed antibody titers against H1 and H3 viruses although H3 seropositivity was more than H1 antibody. This result was similar to other studies representing H3 as one of the predominant subtypes in *Anseriformes* order (24, 25). In this study, seropositivity of ducks against H1N1 and H3N2 were 31.25% and 81.25%, respectively. These results are in accordance with the report of Barbic *et al* (26) in domestic ducks of Croatia. They recorded a positive HI antibody for A/New Caledonia/20/99 (H1N1) and A/Panama /2007/99 (H3N2) viruses in 40.6% and 90.2% of examined sera. Seropositivity of pigeons (*Columbiformes* order) against H3 subtype could be important because they are not routine captive birds of zoos and parks but their population is dominant bird species in some cities such as Tehran. Some tested sera had seropositivity against influenza H1 and H3 viruses while influenza A genome was not detected in any of the cloacal samples. As HI assay is able to detect haemagglutination antibodies as soon as 2 weeks to 1 year post-infection, the results of this study indicate that seropositive captive birds were infected during recent year with H1 and H3 virus strain. Although, the antigens used in HI assay were human viruses, the possibility of cross reaction should be considered. Since the sampling locations had unique niches due to the variety of influenza virus hosts, exposure to H1 and H3 viruses from another origin, including free-flying birds, could have given positive results in the HI test.

**Acknowledgements**

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**References**

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